

Antioxidant and Antibacterial Activities of *Sagina Procumbens*

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Abstract

Sagina Procumbens, a traditional medicinal plant, was examined for its antioxidant activity with the 1, 1-diphenyl-2-picrylhydrazyl (DPPH) assay, its total flavonoid levels with the Folin-Ciocalteu method, and its total phenolics with the Folin-Ciocalteu method. According to the findings of the study, the ethanolic extract of *Sagina Procumbens* leaf has a percentage of antioxidant activity (AA%) of 86.61, which was shown to be greater when compared to other extracts of buds and stem. In a similar vein, the methanolic extract of the stem has a high flavonoid concentration, whereas the phenolic level of the ethanolic extract of the bud is significantly higher. Antimicrobial activity exhibited by *S. procumbens* has a very significant zone of inhibition against gram-positive as well as gram-negative bacterial strains. The findings of this study indicated that plants are excellent sources of natural antioxidants, and that both ethanolic and methanolic extracts of plants contain a sizeable quantity of antioxidant chemicals.

Keywords: *Sagina Procumbens*, extraction in different solvents, antioxidant activity, Total phenolic contents, Total flavonoid contents, Antimicrobial activity.

1. Introduction

Bioactive chemicals have traditionally been derived from plants. The wide availability of chemicals in a natural product like plant extracts opens up endless doors for the discovery of new medicines. Antibacterial drug resistance is a growing problem due to the rapid evolution of germs that cause disease (Al-Dhabi *et al.*, 2015, Al-Dhabi *et al.*, 2016, Barathikannan *et al.*, 2016). That's why scientists from the biological, chemical, and pharmaceutical fields are constantly on the lookout for undiscovered chemicals in the natural world. More and more people are paying attention to medicinal plants because of the wide range of beneficial metabolites found in them (Bonjar and Farrokhi, 2004, Feiginet *et al.*, 2017, Vetrinuruganet *et al.*, 2017).

In the past, people relied on plants as a form of alternative medicine. Nowadays, many medications are made from plants, and hence have no or little adverse effects. Plants are a source for several compounds. Chemicals like this are crucial in the production of pharmaceuticals and personal care products. Plant-based chemicals have long been recognized for their use in the pharmaceutical sector. Many plants have yet to be identified or analyzed for their phytochemical constituents (Jardimet *et al.*, 1998).

Roots, stems, flowers, and leaves, all of which are a part of medicinal plants, all have their own unique set of antibacterial qualities (Elangoet *et al.*, 2016, Fowsiyat *et al.*, 2016, Glorybalet *et al.*, 2015, Harithaet *et al.*, 2016). In traditional medicine, it serves many purposes, including as a pest deterrent, blood thinner, fungal and bacterial killer, and anticoagulant (Helanet *et al.*, 2016, Ilavenilet *et al.*, 2017, Park *et al.*, 2017). This restorative herb has been shown to speed up hair growth and aid in wound healing (Parket *et al.*, 2017). Many nations have turned to herbal remedies to fill their healthcare voids (Awe and Omojalasola, 2003, Surendraet *et al.*, 2017, Roopanet *et al.*, 2019). Activities, such as antibacterial ones, were demonstrated by medicinal plants (Abo *et al.*, 1999).

Validating what is known about medicinal plants in a given region requires both understanding of traditional medical systems and the collaborative efforts of traditional healers and modern scientists (Hamill *et al.*, 2000). Even though traditional medicine is widely utilized in underdeveloped and emerging nations, very little is known about the biological role and chemical makeup of medicinal plants (Tabutiet *et al.*, 2003).

Common names for *Sagina Procumbens* include "procumbent pearlwort," "birdeye pearlwort," and "matted pearlwort," all of which refer to members of the same pink family of blooming plants. It's a perennial plant that grows in mats or clumps of

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smooth, green leaves, often confusingly resembling a mossy area. The linear leaves can grow to be about 2 cm in length. A single flower forms the inflorescence, and it has four or five sepals and four or five tiny white petals (or none at all). The word "sagina" is derived from the Latin word for "fodder," and the genus *Sagina* was originally named after the spurrey plant, which is now classified as its own genus, *Spergula*. What we mean when we say "procumbens" is that we are lying flat on the ground or inching forwards (Botanical Society of Britain and Ireland 2015). This plant can be found in many different parts of the world, but it is very common in Asia. This plant has not been validated for its antibacterial potential, despite the fact that numerous medicinal herbs have been studied for this and other biological functions (Balandrin *et al.*, 1985). *Sagina Procumbens* was thought to have several chemicals with biological activity. The study set out to determine whether or not *Sagina Procumbens* has any useful biological properties.

2. Material and Method

Collection of Plant Sample

The specimen was taken from the MianIshaq Nurseries, and identified there. Once the stem, leaves, and buds were extracted, they were air dried at room temperature in the shade, then milled into a powder for further extraction using various solvents.

Extraction into Different solvents for Biological Activities

To make a powder, we washed, dried in the shade, and then pulverized the plant pieces we had collected. Methanol, ethanol, acetone, and n-Hexane were the solvents of choice for extracting the plant material. The plant powder was mixed with the appropriate solvent at a 1:10 (w/v) ratio in a clean flask. The flasks were incubated at 37 degrees Celsius and 10,000 rpm for four days. Then, it is filtered through No. 1 Whatman's filter paper, condensed using a rotary evaporator, and chilled to 4 degrees Celsius so that additional analysis can be performed (Den *et al.*, 2020).

Total Phenolic Content

The total phenolic content was determined by using the Folin-Ciocalteu reagent and the same methodology published by Ainsworth and Gillespie (2007), with some minor adjustments. The sample extract (100 L) was mixed with the diluted FolinCiocalteu reagent (200 L) (10%) and vigorously vortexed. The absorbance at 765 nm was then measured after each sample was treated with Na₂CO₃ (800 L of 700 mM) and left to stand at room temperature for 2 hours. When converting to GAE from mg/g, gallic acid was employed as the benchmark.

Total Flavonoids Content

The method for measuring total flavonoid content was adapted from that published by Anjum *et al.*, (2013). After taking a 1mL sample extract in a test tube, 3mL of 5% NaNO₂ was added, followed by 0.6mL of AlCl₃ (10%) after 5 minutes and 2mL of NaOH (1M). The 510 nm absorbance reading was recorded. Catechin is the benchmark (in milligrams per gram of dry substance).

DPPH Free Radical Activity

A sample (50 L) was diluted in 5 mL of DPPH methanol solution (0.004%). After incubation for 30 minutes at room temperature, the absorbance was measured at 517 nm.

$$I\% = (A \text{ blank} - A \text{ sample} / A \text{ blank}) * 100$$

In which A_{blank} is the absorbance of DPPH solution and A_{sample} being the absorbance of sample (Bozin *et al.*, 2006).

Antimicrobial Activities

Extracts from plants were tested for their ability to kill off certain strains of bacteria, including Gram-negative *E. coli* and Gram-positive *Bacillus subtilis* (CLSI, 2007).

Anti-Bacterial Assay by Disc Diffusion Method

The nutrient agar solution (28.08 g/L) was autoclaved at 121 degrees Celsius for 15 minutes. When the media reached room temperature after being sterilized, various bacterial species were introduced. Positive control was ampicillin, and negative control was distilled water. Solution was poured onto sterile discs prepared from Whatsmann filter paper No. 1, which were then placed on solidified media poured into petri dishes. Millimeter-scale readings from a zone reader confirmed the presence of a distinct zone of inhibition around the antibacterial extract (Banerjee *et al.*, 2014).

Statistical Evaluation

Minitab version 10 was used for the two-way analysis of variance and the least significant difference between the solvents.

3. Results and Discussion

Total Phenolic Contents

Phenols are important compounds that are widely distributed across plant kingdom. There may be a connection between the total phenolic contents and total antioxidant activity of the plant extracts, and this connection might be shown by qualitative and quantitative research into phenolic content in plants (Demain *et al.* 2009). Polyphenolic compounds have recently been shown to have an important role in warding off degenerative diseases like cardiovascular disease, cancer, and neurodegenerative disorders. Diets high in polyphenols have been shown to have

beneficial effects on health, and the processes behind this are becoming clearer thanks to the antioxidant properties of these compounds.

Numerous therapeutic plants have high concentrations of polyphenolic substances like phenolic acids, tannins, and flavonoids (Donget *al.*, 2009). Phenolic acids are extensively distributed secondary aromatic plant metabolites. These phenolic acids can be distinguished from one another by their hydroxycinnamic and hydroxybenzoic structures, both of which are naturally occurring. The antioxidant and health benefits of foods have been linked to their phenolic acid content, as well as their color and flavor (Magaet *al.*, 2008).

Figure 1 shows that the total phenolic content of the ethanolic extract of *Sagina Procumbens* leaf is much higher than that of the other extracts tested (65.181.04). Leaf extract by acetone showed similarly low phenolic levels. The polarity of the solvent we utilized for extraction explained the variation in phenolic content. When compared to other solvents, ethanol was discovered to be the most polar. To put it another way, the phenolic content of *Sagina Procumbens* leaves is highest in ethanolic extracts and lowest in acetone extracts (31.621.07 mg/g). The total phenolic content of the methanolic extract of *Sagina Procumbens* stem is also quite high (45.680.71). The phenolic content of leaf extract prepared with acetone was likewise found to be quite low.

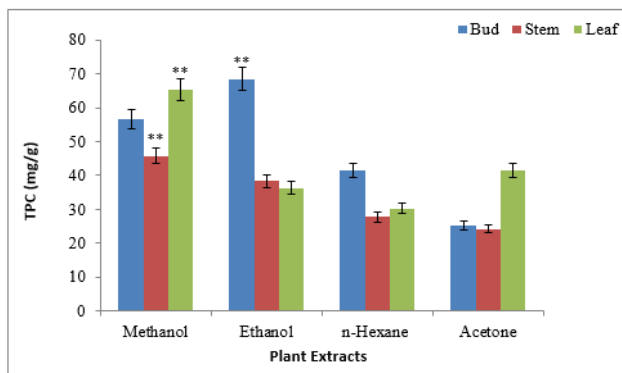


Figure 1. Total phenolic contents of Saginaprocumbens

When compared to other extracts, the ethanolic extract of *Sagina Procumbens* bud has a very high total phenolic content (68.380.99). The phenolic content of bud extract prepared with acetone was also found to be low (25.131.05). The ethanolic extract of *Sagina Procumbens* buds had a higher total phenolic content than the stems and leaves. Because this plant species has not been studied, it is obvious that bud extract of ethanol contains more phenolic compounds than stem and leaves.

Total Flavonoid Contents

Flavonoids have a variety of pharmacological and biological functions, including metal chelation,

scavenging of reactive oxygen species and chain-breaking antioxidants, reduction of agents, quenching of singlet oxygen, and other related actions (Naseeret *al.*, 2014). Several illnesses of the neurological system and inflammation are treated with antioxidants (Babu et *al.*, 2011).

Water-soluble complexes like flavonoids and lipid-soluble complexes like phenolic acids are two types of phytochemicals. Total flavonoid content was estimated using a method that has been utilized in numerous previous studies (Ghimerrayet *al.*, 2009).

Figure 2 shows that the total flavonoid content of *Sagina Procumbens* stem extracts prepared using methanol and n-hexane are, respectively, 78.530.84 and 31.020.73.

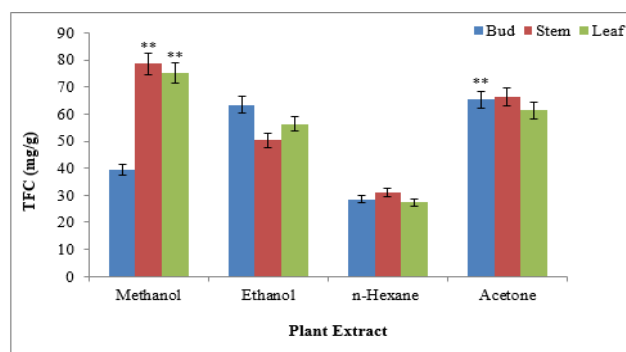


Figure 2. Total Flavonoid Contents of *Sagina Procumbens*

Flavonoid concentration was found to be much higher (66.351.10) in acetone extract of leaf than in phenolic content, perhaps due to the polarity of the solvents used. To a similar extent, the flavonoid content of *Sagina Procumbens* leaves extracted with methanol is higher than that of leaves extracted with n-hexane (27.440.96 mg/g). *Sagina Procumbens* bud acetone extract had the highest total flavonoid concentration (65.270.84 mg/g) when compared to other extracts. N-hexane bud extract, on the other hand, is not very rich in flavonoids. Flavonoids are the polyphenolic substances that have shown promise for improving human health; they have been shown to have antiviral, anti-inflammatory, anticancer, antithrombotic, and anti-oxidative actions (Buhler and Miranda, 2000).

DPPH Radical Scavenging Assay

We used the compound's ability to scavenge DPPH free radicals to evaluate its antioxidant activity in a series of synthesized compounds. As a free radical, 1, 1-diphenyl-2-picryl-hydrazyl is violet in color but turns yellow upon reduction. It is soluble in methanol. Antioxidant activity is proportional to an antioxidant's ability to quench the DPPH radical. The percentage of the DPPH radical that is inhibited provides a great explanation of antioxidant efficacy. The presence of antioxidant compounds is confirmed by the radical scavenging activity of ethyl acetate extracts and their ability to donate electrons or hydrogen atoms.

When a 2, 2-diphenyl-1-1-hydrazyl reagent accepts an electron, the molecule becomes diamagnetic and also functions as a free radical. The DPPH assay was proven to be a quick and inexpensive way to determine an antioxidant's effectiveness. The DPPH technique was described by Souri et al.(2008). The byproducts of photosynthesis and photorespiration are reactive oxygen species. When added to samples, DPPH takes a few seconds to lose its purple hue, and after 10 minutes in incubation, its once-vivid hue fades to a pastel yellow.

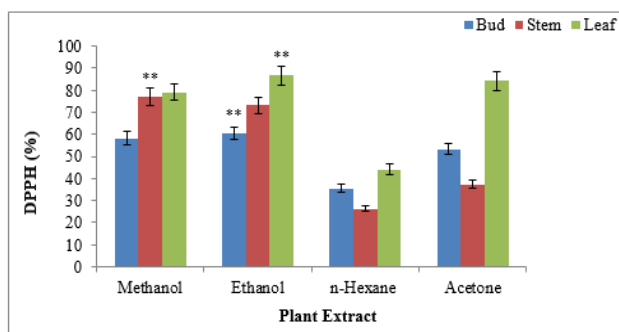


Figure 3. DPPH activity of Sagina Procumbens

Figure 3 shows that the methanolic extract of Sagina procumbens's stem has the highest DPPH activity (77.121.02%), while the n- hexane extract has the lowest (26.30.67%). Due to the polarity of the solvents, it was also observed that the ethanol extract of the leaf exhibited a considerably high (86.611.07) percentage of DPPH activity. The percentage of DPPH activity in n-hexane extract is lowest (44.211.20). The DPPH activity of ethanolic extracts of Saginaprocumbens buds is comparable to that of n-hexane extracts, whereas the DPPH activity of n-hexane extracts is the lowest (35.460.94%). In addition, the DPPH concentration of ethanolic extract of Sagina procumbens leaf was found to be substantially higher than that of bud and stem extracts obtained using different solvents.

Antimicrobial Activity of Sagina Procumbens

Antimicrobial activity was tested in Sagina Procumbens extracts. Some extracts had a wide range of activity, as evidenced by a distinct zone of inhibition,

Table 1. Comparative analysis of antibacterial activity of different extracts of stem leaf and bud of Saginaprocumbens against gram positive (B.subtilis) and gram negative (E.coli) bacterial strainwhile others were ineffective against the strain at hand.

Sample extracts	Zone of inhibition (mm)					
	<i>E.coli</i>			<i>B.subtilis</i>		
	Stem	Leaf	Bud	Stem	Leaf	Bud
Methanol	16.00±0.77 ^A	14.00±0.61 ^B	12.00±0.54 ^C	14.00±0.61 ^B	16.00±0.67 ^A	16.00±0.67 ^A
Ethanol	14.00±0.61 ^B	12.00±0.54 ^C	12.00±0.54 ^C	16.00±0.77 ^A	12.00±0.54 ^C	16.00±0.67 ^A
n-Hexane	12.00±0.54 ^C	12.00±0.54 ^C	14.00±0.61 ^B	14.00±0.61 ^B	12.00±0.54 ^C	12.00±0.54 ^C
Acetone	16.00±0.77 ^A	14.00±0.61 ^B	14.00±0.61 ^B	14.00±0.61 ^B	14.00±0.61 ^B	12.00±0.54 ^C
Ampicillin (positive control)	22.00±0.99 ^A	20.00±0.99 ^A	22.00±0.99 ^A	24.00±1.23 ^A	24.00±1.23 ^A	24.00±1.23 ^A

If the results are negative, it means that the plant either did not contain any active compound, or if it did, the concentration was so low that the compound was ineffective.

In comparison to the stem extract, the leaf and bud methanolic extracts of Sagina Procumbens showed significantly higher antibacterial activity (16.000.67) against Gram positive bacteria (B. subtilis). Sagina Procumbens leaf and bud extract are less effective against Gram-negative bacteria (E. coli) than the plant's stem extract (16.000.77). The results were measured against those obtained when using ampicillin (an antibiotic). Stem and bud extracts of Sagina Procumbens have excellent antibacterial activity (16.000.77) against B.subtilis(Gram positive bacteria), whereas stem extracts had good antibacterial activity (14.000.61) against gram negative bacteria (E.coli).

Table 1 displays data indicating that, back Against B.subtilis (Gram positive bacteria), hexane extract of Sagina Procumbens stem has strong (14.000.61) antibacterial activity, while bud extract has high antibacterial activity (14.000.61) against gram negative bacteria (E.coli). Sagina Procumbens stem, leaf, and bud acetone extracts have antibacterial action against B.subtilis(Gram positive bacteria) (14.000.61), whereas stemextract has considerable antibacterial activity against gram negative bacteria (E.coli) (16.000.77). It is evident from the foregoing that the zone of inhibition of antibacterial activity against B.subtilisas is significantly higher for the methanolic and ethanolic extract of the stem of Sagina Procumbens than it is for E.coli. Standard antibiotic treatment was used as a benchmark for all of these findings (ampicilin).

Damage to the bacterial cell wall, membrane, enzyme inhibition or metabolic inactivation, and bacterial nucleic acids molecules are the primary mechanisms by which chemicals of any origin exert their antibacterial effect (Alsughayer et al., 2011). Chemically interfering with the bacterial cell wall or membrane, or entering the cell and chemically altering the proteins or nucleic acids, are all effects of antibacterial chemicals' various pharmacologically active functional groups, which have electron withdrawing or electron giving capacities. Cell growth arrest or death is the cumulative effect of such changes (Kumaret al., 2010).

Some of these chemicals are structural analogues of metabolic intermediates and function as enzyme inhibitors or modifiers of enzyme bases involved in essential catalytic reactions for bacterial cell survival (Cossuet al., 2012). Some sulfonamides, such as sulfanilamide, are structurally similar to tetrahydrofolate, a bacterial precursor to folic acid synthesis; however, because of structural differences, these sulfonamides are unable to enter the active site of the tetrahydrofolate synthase enzyme, thereby halting the growth of the bacteria. Distinct chemicals with possible antibacterial effects have had several different mechanisms reported (Kucukguzel, et al., 2002). *Sagina Procumbens* has been discovered to have strong potential against pathogens, despite the fact that it has not been the subject of extensive prior investigation.

Conclusion

Plants with medicinal properties have been utilized for ages to heal human illness because of the substances they contain. Flavonoids are a naturally occurring chemical with a wide range of pharmacological effects, including those of an anti-inflammatory, anti-oxidant, and diuretic. Researchers are now able to explore the photochemical and antimicrobial activity of many herbs and medicinal plants thanks to antibiotic resistance available from these sources. Before this experiment, *Sagina Procumbens* wasn't given much of a second thought, but now that we know it has microbiological resistance against infections and active potential of antioxidant chemicals and is getting a lot more attention.

References

- [1]. Abo, A. M., & Gray, P. R. (1999). A 1.5-V, 10-bit, 14.3-MS/s CMOS pipeline analog-to-digital converter. *IEEE Journal of Solid-State Circuits*, 34(5), 599-606.
- [2]. Ainsworth, E. A., & Gillespie, K. M. (2007). Estimation of total phenolic content and other oxidation substrates in plant tissues using Folin-Ciocalteu reagent. *Nature protocols*, 2(4), 875-877.
- [3]. Al-Dhabi, N. A., Arasu, M. V., Park, C. H., & Park, S. U. (2015). An up-to-date review of rutin and its biological and pharmacological activities. *EXCLI journal*, 14, 59.
- [4]. Al-Dhabi, N. A., Esmail, G. A., Duraipandiyam, V., Arasu, M. V., & Salem-Bekhit, M. M. (2016). Isolation, identification and screening of antimicrobial thermophilic *Streptomyces* sp. Al-Dhabi-1 isolated from Tharban hot spring, Saudi Arabia. *Extremophiles*, 20(1), 79-90.
- [5]. Alsughayer, A., Elassar, A. Z. A., Mustafa, S., & Al Sagheer, F. (2011). Synthesis, structure analysis and antibacterial activity of new potent sulfonamide derivatives. *Journal of Biomaterials and Nanobiotechnology*, 2(02), 143.
- [6]. Anjum, N. A., Singh, N., Singh, M. K., Shah, Z. A., Duarte, A. C., Pereira, E., & Ahmad, I. (2013). Single-bilayer graphene oxide sheet tolerance and glutathione redox system significance assessment in faba bean (*Vicia faba* L.). *Journal of nanoparticle research*, 15(7), 1770.
- [7]. Awe, S., & Omojalasola, P. F. (2003). Antibacterial screening of three medicinal plants used for diarrhea treatment in Ilorin, Nigeria. *Nig. J. Pure and Appl. Sci*, 1, 1375-1379.
- [8]. Babu, M. M., van der Lee, R., de Groot, N. S., & Gsponer, J. (2011). Intrinsically disordered proteins: regulation and disease. *Current opinion in structural biology*, 21(3), 432-440.
- [9]. Balandrin, M. F., Klocke, J. A., Wurtele, E. S., & Bollinger, W. H. (1985). Natural plant chemicals: sources of industrial and medicinal materials. *Science*, 228(4704), 1154-1160.
- [10]. Banerjee, S., Carlin, B. P., & Gelfand, A. E. (2014). Hierarchical modeling and analysis for spatial data. CRC press.
- [11]. Barathikannan, K., Venkatadri, B., Khusro, A., Al-Dhabi, N. A., Agastian, P., Arasu, M. V., & Kim, Y. O. (2016). Chemical analysis of *Punicagranatum* fruit peel and its in vitro and in vivo biological properties. *BMC complementary and alternative medicine*, 16(1), 264.
- [12]. Bonjar, G. S., & Farrokhi, P. R. (2004). Anti-bacillus activity of some plants used in traditional medicine of Iran. *Nigerian Journal of Natural Products and Medicine*, 8, 34-39.
- [13]. Bozin, B., Mimica-Dukic, N., Simin, N., & Anackov, G. (2006). Characterization of the volatile composition of essential oils of some Lamiaceae spices and the antimicrobial and antioxidant activities of the entire oils. *Journal of agricultural and food chemistry*, 54(5), 1822-1828.
- [14]. Brand-Williams, O., & Kiska, T. (1999). NAACP condemns fall TV: boycott, lawsuit vowed over lack of racial diversity. *Detroit News*.
- [15]. Cossu, G., Khalid, A. M., Choudhury, P., Corsini, R., & Ciaramella, E. (2012). 3.4 Gbit/s visible optical wireless transmission based on RGB LED. *Optics express*, 20(26), B501-B506.
- [16]. Demain, A. L., & Vaishnav, P. (2009). Production of recombinant proteins by microbes and higher organisms. *Biotechnology advances*, 27(3), 297-306.
- [17]. Deng, J., Dong, W., Socher, R., Li, L. J., Li, K., & Fei-Fei, L. (2009, June). Imagenet: A large-scale hierarchical image database. In 2009 IEEE conference on computer vision and pattern recognition (pp. 248-255). Ieee.
- [18]. Elango, G., & Roopan, S. M. (2016). Efficacy of SnO2 nanoparticles toward photocatalytic degradation of methylene blue dye. *Journal of Photochemistry and Photobiology B: Biology*, 155, 34-38.
- [19]. Feigin, V. L., Abajobir, A. A., Abate, K. H., Abd-Allah, F., Abdulle, A. M., Abera, S. F., ... & Nguyen, G. (2017). Global, regional, and national burden of neurological disorders during 1990-2015: a systematic analysis for the Global

- Burden of Disease Study 2015. *The Lancet Neurology*, 16(11), 877-897.
- [20]. Fowsiya, J., Madhumitha, G., Al-Dhabi, N. A., & Arasu, M. V. (2016). Photocatalytic degradation of Congo red using *Carissa edulis* extract capped zinc oxide nanoparticles. *Journal of Photochemistry and Photobiology B: Biology*, 162, 395-401.
- [21]. Ghimeray, A. K., Jin, C. W., Ghimire, B. K., & Cho, D. H. (2009). Antioxidant activity and quantitative estimation of azadirachtin and nimbin in *Azadirachta Indica* A. Juss grown in foothills of Nepal. *African Journal of Biotechnology*, 8(13).
- [22]. Glorybai, L., Arasu, M. V., Al-Dhabi, N. A., & Agastian, P. (2015). Some biological activities of *Epaltesdivaricata* L-an in vitro study. *Annals of Clinical Microbiology and Antimicrobials*, 14(1), 18.
- [23]. Hamill, T. M., & Snyder, C. (2000). A hybrid ensemble Kalman filter-3D variational analysis scheme. *Monthly Weather Review*, 128(8), 2905-2919.
- [24]. Haritha, E., Roopan, S. M., Madhavi, G., Elango, G., Al-Dhabi, N. A., & Arasu, M. V. (2016). Green chemical approach towards the synthesis of SnO₂ NPs in argument with photocatalytic degradation of diazo dye and its kinetic studies. *Journal of Photochemistry and Photobiology B: Biology*, 162, 441-447.
- [25]. Helan, V., Prince, J. J., Al-Dhabi, N. A., Arasu, M. V., Ayeshamariam, A., Madhumitha, G., ... & Jayachandran, M. (2016). Neem leaves mediated preparation of NiO nanoparticles and its magnetization, coercivity and antibacterial analysis. *Results in physics*, 6, 712-718.
- [26]. Ilavenil, S., Kim, D. H., Srigopalram, S., Kuppasamy, P., Arasu, M. V., Lee, K. D., ... & Choi, K. C. (2017). Ferulic acid in *Lolium multiflorum* inhibits adipogenesis in 3T3-L1 cells and reduced high-fat-diet-induced obesity in Swiss albino mice via regulating p38MAPK and p44/42 signal pathways. *Journal of Functional Foods*, 37, 293-302.
- [27]. Jardim, C. M., Jham, G. N., Dhingra, O. D., & Freire, M. M. (2008). Composition and antifungal activity of the essential oil of the Brazilian *Chenopodium ambrosioides* L. *Journal of Chemical Ecology*, 34(9), 1213-1218.
- [28]. Küçükgülzel, Ş. G., Oruç, E. E., Rollas, S., Şahin, F., & Özbek, A. (2002). Synthesis, characterisation and biological activity of novel 4-thiazolidinones, 1, 3, 4-oxadiazoles and some related compounds. *European Journal of Medicinal Chemistry*, 37(3), 197-206.
- [29]. Kumar, V., Yadav, C. S., Singh, S., Goel, S., Ahmed, R. S., Gupta, S., & Banerjee, B. D. (2010). CYP 1A1 polymorphism and organochlorine pesticides levels in the etiology of prostate cancer. *Chemosphere*, 81(4), 464-468.
- [30]. Maga, G., Falchi, F., Garbelli, A., Belfiore, A., Witvrouw, M., Manetti, F., & Botta, M. (2008). Pharmacophore modeling and molecular docking led to the discovery of inhibitors of human immunodeficiency virus-1 replication targeting the human cellular aspartic acid-glutamic acid-alanine-aspartic acid box polypeptide 3. *Journal of medicinal chemistry*, 51(21), 6635-6638.
- [31]. Miranda, C. L., Stevens, J. F., Ivanov, V., McCall, M., Frei, B., Deinzer, M. L., & Buhler, D. R. (2000). Antioxidant and prooxidant actions of prenylated and nonprenylated chalcones and flavanones in vitro. *Journal of agricultural and food chemistry*, 48(9), 3876-3884.
- [32]. Naseer, N., Noori, F. M., Qureshi, N. K., & Hong, K. S. (2016). Determining optimal feature-combination for LDA classification of functional near-infrared spectroscopy signals in brain-computer interface application. *Frontiers in human neuroscience*, 10, 237.
- [33]. Park, S., Yoon, Y. M., Han, S. K., Kim, D., & Kim, H. (2017). Effect of hydrothermal pre-treatment (HTP) on poultry slaughterhouse waste (PSW) sludge for the enhancement of the solubilization, physical properties, and biogas production through anaerobic digestion. *Waste Management*, 64, 327-332.
- [34]. Roopan, S. M., Priya, D. D., Shanavas, S., Acevedo, R., Al-Dhabi, N. A., & Arasu, M. V. (2019). CuO/C nanocomposite: Synthesis and optimization using sucrose as carbon source and its antifungal activity. *Materials Science and Engineering: C*, 101, 404-414.
- [35]. Souri, M., Azarmanesh, M. N., Sani, E. A., Nasseri, M., & Farhadi, K. (2008). An analytical study of resistive oxygen gas sensors. *Journal of Physics: Condensed Matter*, 20(14), 145204.
- [36]. Surendra, K. C., Olivier, R., Tomberlin, J. K., Jha, R., & Khanal, S. K. (2016). Bioconversion of organic wastes into biodiesel and animal feed via insect farming. *Renewable energy*, 98, 197-202.
- [37]. Tabuti, J. R., Dhillon, S. S., & Lye, K. A. (2003). Ethnoveterinary medicines for cattle (*Bos indicus*) in Bulamogicounty, Uganda: plant species and mode of use. *Journal of Ethnopharmacology*, 88(2-3), 279-286.
- [38]. Vetrinmurugan, E., Brindha, K., Elango, L., & Ndwandwe, O. M. (2017). Human exposure risk to heavy metals through groundwater used for drinking in an intensively irrigated river delta. *Applied Water Science*, 7(6), 3267-3280.