

Original Research Article

Putative amniotic fluid stem (AFS) cells express transcription factor *Oct-4* in goat (*Capra aegagrus hircus*)

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Abstract

The current study was carried out to isolate, culture, characterize and cryopreserve the putative amniotic fluid stem (AFS) cells in goat. Putative AFS cells were cultured without feeder cells, in DMEM containing 10% FBS, 1% penicillin/streptomycin and 1% L-glutamine in 5% CO₂ in humidified air at 39.1±0.5°C. After 6 days of culture different morphologies of cells were observed. Most of the cells started anchorage-dependent growth after day 6 of the culture. In order to check their pluripotency properties when the cells were subjected to RT-PCR a strong positive expression of *Oct-4* was observed. Using species-specific primers, a PCR amplicon of 156 bp was observed for *Oct-4*. When the cells were cryopreserved in DMSO containing freezing media, >70% of the cells were viable. These results may contribute towards establishing non-embryonic pluripotent stem cells for various therapeutic and reproductive biotechnological applications in this species.

Keywords: Goat, amniotic fluid, stem cells, characterization and RT-PCR.

1. Introduction

Goat an important species of livestock for India represents goat biodiversity in the form of breeds and strain. The domestication of animals is being carried out from Neolithic times along with the cultivation of cereals. First sheep and goats, second cattle and pigs, and finally draft animals such as horses and asses were domesticated. They contribute greatly to the agrarian economy, especially in areas where crop and dairy farming are not economical, and play an important role in the livelihood of a large proportion of small and marginal farmers and landless laborers. Among the domesticated livestock species, there are over 30 breeds of cattle, 10 breeds of river buffaloes, 42 breeds of sheep, 20 breeds of goats, 8 breeds of camel, 7 breeds of horse and 18 of the indigenous poultry. India's vast genetics resources in the goat are reflected by the availability of 20 breeds of goat. Small ruminants especially goat contribute to the livelihoods of millions of rural poor in most of the developing countries of the Asia and Africa, However, introduction of exotic breeds, natural calamities and change in farming system have been resulted decline in pure breed population and in dilution of the genetics merit.

The productivity of Indian goat is low, yet considering the nutritional and physical environmental conditions under which they reared it cannot be considered inefficient. Major reasons for this low productivity are inadequate grazing resources, disease problems and serious lack of organized efforts for genetic improvement. There is little selection of rams and bucks used for breeding, and much inter-mating among neighboring breeds takes place. The traditional breeding program involving selections based on production performance have contributed significantly to improvement of production performance of these goats. However, these techniques require long duration to improve to a particular trait. Therefore, researchers are attempting to improve goat reproduction through innovative approaches using various reproductive technologies. Coordinated systems of reproductive management have been developed based on a thorough understanding of the endocrine, cellular and molecular factors controlling ovarian and uterine functions. Reproductive endocrinological interventions have contributed to improvements in goat productivity, particularly through increased embryo production and the birth of the goat through embryo transfer technology. Notable reports have presented a comprehensive account of the information on

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reproductive endocrinological advances, including ovarian follicular dynamics, knowledge of which may lead to better synchronization (as well as embryo production and transfer) in goats.

The productive and reproductive efficiency of goat is poor due to lower genetic potential which has not been studied in relation to functional genomics. Stem cells represent an ideal tool to study embryogenesis under in vitro conditions, particularly the genes involved in the functional development. Moreover, availability of goat stem cells could facilitate development of efficient methods for somatic cell cloning, transgenics, and gene regulation in this species. As far as the goat amniotic fluid stem cells are concerned, there is very limited information available on these lines. To the best of our knowledge, no work has been carried out on scanning electron microscopy for morphological studies of these cells. Looking at scarcity of the information and usefulness of these cells the current study was designed.

2. Materials and Methods

2.1 Chemicals and media

All chemicals i.e. reagents, culture media and antibiotics used during the study were of cell culture grade, obtained from Hi Media Laboratories (Mumbai, India) unless otherwise indicated. Trizol was from Invitrogen (USA). Disposable 35 mm x10 mm cell culture Petri dishes, 6 well tissue culture plates, and centrifuge tubes were procured from Tarsons Products Pvt. Ltd. (Kolkata, India). Membrane filters were from Advanced Micro-devices (Ambala, India). The primers were got synthesized from GenxBio (India). The culture media were reconstituted freshly as per manufacturers' instructions and filter-sterilized (0.22µm) prior to use.

2.2 Transportation and collection of sample

Goat amnion was obtained from a nearby abattoir, washed 2-3 times with isotonic saline fortified with 1% of penicillin/streptomycin and transported to the laboratory in a thermally insulated ice box within 5 hour. Uterine cut, fetus and membranes were located and AF was aspirated aseptically with the help of 20 ml syringe fitted with 16 gauge hypodermic needle. Twenty milliliters of AF was collected in Oakridge tubes. The appearance (fluid without cells, blood-cell-free, blood-cell, bloody or brown-coloured fluid), of fluid volume which was collected were observed and carefully documented.

2.3 Isolation and culture of putative AFS cells

The putative AFS cells were separated by centrifugation. Amniotic fluid was centrifuged at 3000g for 10 min and washed twice with phosphate buffered saline (PBS) and this procedure was repeated two to

three times. During centrifugation a layer was formed at bottom of the tube and a buffycoat formed in which blood cells were seen.

2.4 Culturing of putative AFS cells

The cells were ready to culture. The cells were seeded at density of 10^3 cells / cm^2 in a 6 well culture plates containing cell culture medium (DMEM) supplemented with 10% FBS (fetal bovine serum) and 1% Penicillin / Streptomycin / Ampicillin. Cultured plates were incubated in humidified CO₂ Incubator (Lark, China) at $39.1 \pm 0.5^\circ\text{C}$ in presence of 5% CO₂ in humidified air (Dev et al., 2012). The cells were allowed to grow and were subcultured by passaging after achieving >80% confluency.

2.5 Oct-4 expression by Reverse Transcriptase PCR

2.5.1 mRNA synthesis

The method proposed (Hummon et al; 2007) with minor modifications was followed for extraction of total cellular RNA. RNA was extracted from approximately 0.6×10^7 cells using the Trizol agent. Procedure adopted was as follows: 300 µl sample of putative amniotic fluid stem cells was taken and 700 µl of Trizol reagent was added in an eppendorf tube and mixed properly. 200 µl chloroform was added, gently mixed for 5 minutes. Centrifugation was carried out at 12000g for 15 min. Two separate layers i.e. aqueous and organic were formed. Aqueous layer was carefully taken and transferred to a fresh eppendorf tube. To it added 198 µl chloroform and 8 µl isoamyl alcohol (24:1), mixed and centrifuged at 12000g for 15 min. The supernatant obtained was collected. To it 500 µl isopropyl alcohol was added then again centrifuged at 12000g for 5 min. Upper layer was carefully removed and 700 µl absolute alcohol was added and centrifuged at 12000g for 10 min. Supernatant was discarded and 70 µl RNAase free DEPC-treated water was added for removing RNAases. RNA was prepared then run on 0.8% agarose gel using agarose gel electrophoresis.

2.5.2 cDNA formation

The cDNA was synthesized by reverse transcription of mRNA purified from the putative AFS cells. The reaction mixture comprised of total cellular 5 ng RNA, 0.2 µg / µl random hexamer, 7 µg / µl cDNA direct RT, 10 µM / µl AMV reverse transcriptase and 40 U / µl RNase inhibitor in a total volume of 20 µl (one step RT-PCR kit, Novagen). The primer sequences used for Oct-4 was 5'-GTTCTCTTTGGAAAGGTGTTTC-3' and 5'-ACACTCGGACCACGTCTTTC-3'. The reverse transcriptase PCR (RT-PCR) was carried out at 42°C for 60 min. followed by denaturation at 95°C for 8 min. After RT-PCR the gel was run on 1.8% agarose.

2.6 Cryopreservation and thawing of putative AFS cells

The putative AFS cells were cryopreserved using slow freezing method. Cells were harvested in a sterilized tube and then centrifuged at 170 g for 5 min. The supernatant was removed from the tube and the cell pellet was resuspended with freezing medium (50% FBS, 40% DMEM and 10% DMSO) to obtain a final cell density of $1-2 \times 10^6$ cells/mL. Cells were transferred into cryogenic vials. The vials were kept at 4° C for 30 min and maintained at -20° C overnight then were stored in liquid nitrogen (LN₂) for long term preservation. The cryogenic vials were kept in LN₂ for 38 hrs. For thawing, the vials were taken out of the LN₂ and thawed quickly in a 37° C water bath. Vial contents were transferred into a tube containing 2-3 mL of culture medium (antibiotic free), centrifuged at 170 g for 5 min and suspended with an appropriate volume of medium for the dishes. The viability after thawing was also checked as explained below.

2.7 Viability assay of putative AFS cells

For measuring the viability of the cells trypan blue assay was performed. The preparation of 1:1 dilution of the suspension using a 0.4% trypan blue solution was followed. Briefly the cell suspension was loaded in the counting chambers of a hemacytometer with the dilution and observed the cells under compound microscope.

3. Results

3.1 Culturing of putative AFS cells

After collection, all the cells were spherical and variable in sizes.

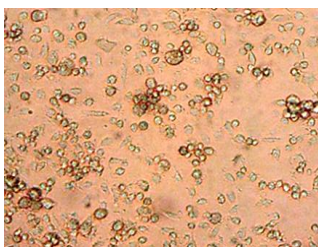


Fig. 1 (a) Putative AFS cell after 48-72 hr of culturing

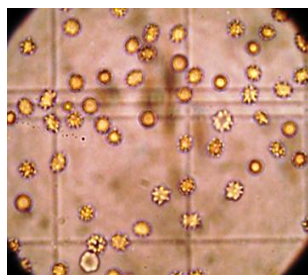


Fig. 1 (b) Different morphologies of amniotic fluid cells after day 6

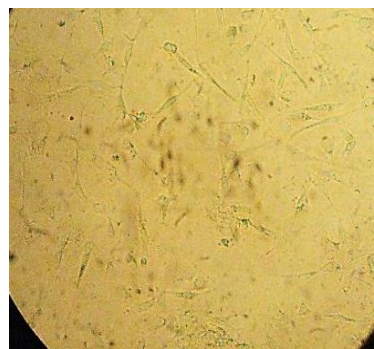


Fig. 1 (c) Fibroblast like cells after day 15 of culturing

No anchorage was observed before 48–72 h (Fig. 1a) of culturing the cells. After day 6, morphologically different cells viz., star shaped (62.7%), spherical without nucleus (1.9%), spherical with nucleus (26.4%), pentagonal (0.4%), and free floating and rounded cells (8.3%) were observed (Fig. 1b). At day 7 of culturing, most of the cells converted into star shaped cells which subsequently started anchoring to surface. At 15th day of culturing the anchorage-dependent cells subsequently gained typical fibroblast like shape and formed a confluent monolayer (Fig. 1c). A few spherical and freely floating cells were also visible. Instead of forming uniform cell monolayer, certain cell clumps were also observed. Initially, the cells reached 70–80% confluence after two weeks. However, the passaged cells exhibited higher growth rate, reaching a 90–100% confluence after day 6 of culturing.

3.2 Viability after cryopreservation of putative AFS cells

After cryopreservation and thawing, the viability assay was performed to check the percentage of viable and dead cells. It was observed that >70% of the cells were viable after trypan blue staining (Fig. 2).

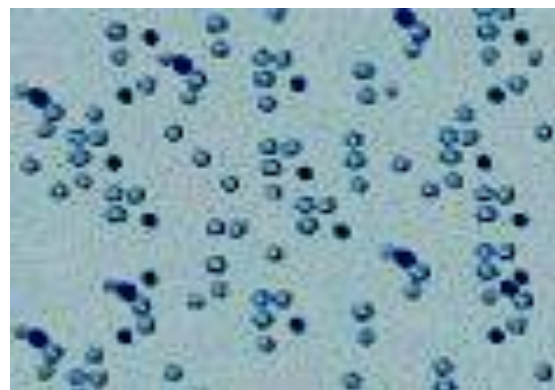


Fig.2 Viability assay: Light blue colour cells are viable cells and dark blue cells are dead cell

3.3 Characterization of putative AFS cells by Oct-4 expression

Agarose gel electrophoresis for analysis of RT-PCR product revealed a 156 bp amplicon of Oct-4 (Fig. 3).

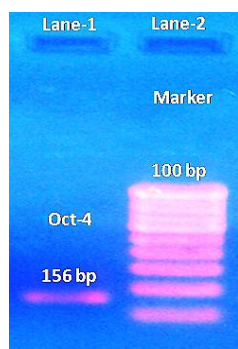


Fig.3 Oct-4 expression in goat putative AFS cells using RT-PCR at day 6 of the culture. Lane 1: 156 bp Amplified product of Oct-4. Lane-2: 100 bp marker.

4. Discussion

Mammalian AF contains diverse cell types representative of three germ layers (Gosden, 1983; Fauza, 2004). Amniotic membrane and AF-derived cells have therefore, attracted a deal of attention globally as an alternative cell source for transplantation and tissue engineering, and as a possible reserve of pluripotent stem cells that may be useful for clinical application in regenerative medicine (Atala, 2006; Parolini et al., 2009; Dobreva et al., 2010) and reproductive biotechnological applications (Zhao and Zheng, 2010). However, the potential of AF stem cells in livestock assisted reproduction and health applications are yet to be exploited. The establishment of pluripotent stem cell lines in domestic species could have great impact in the agricultural as well as in the bio- medical field. Accordingly, the study of the AF stem cells in live-stock species has become a new focus recently (Zhang and Chen, 2008). Efforts are being made to study various types of stem cells (Verma et al., 2007; Dev et al., 2012; Sritanaudomchai et al., 2007) in domestic animals. The present study is a preliminary effort to investigate whether the AFS cells in goat AF can be cultured and exhibit stem cell-like attributes. It has been observed that goat AF cells were able to grow without feeder cells. The choice of the culture medium and conditions chosen to grow goat AF cells are based on reports already established for human AF stem cells (De Coppi et al., 2007). However, the final selection was based on our preliminary observations on the growth of the cells in various combinations of culture media and supplements in buffalo species (Dev et al., 2012). After 3 to 5 days of incubation, the goat AF cells had five different types of morphologically different cells (Fig. 1 a,b). The polygonal or star shaped cells were cultured for prolonged periods (at >10th passage). It was found that these cells transformed into fibroblast-like cells (Fig. 1c). The cells had similar morphology to the AFS cells of buffalo (Dev et al., 2012) and as also reported by Mihiu et al., 2009 where the authors observed the AF stem cells to show morphological features similar to fibroblasts. The goat AF cells were found to have enlarged nuclei compared to adult skin fibroblasts, cumulus cells and granulose cells (data not shown). The AF cells, which were initially round in structure, started anchorage-

dependent growth after day 3 to 5 of culture in vitro. For confirmation whether the cells had pluripotency properties or not, a strong positive expression of Oct-4 was found which indicates that these cells have the ES cell like cell's attributes. However for more confirmative studies regarding their pluripotency, more pluripotency markers need to be studied. After cryopreservation, the cells had good viability which indicates that these cells can be cryopreserved successfully and can be used later on.

In summary, the present study is a preliminary attempt on isolation, culturing and cryopreservation of putative amniotic fluid stem cells in goat. These putative AFS cells can be used in various purposes in future for various investigations. However, use of AFS cells in goat therapeutic and assisted reproductive biotechnology needs further studies.

5.0 Acknowledgement

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References

- Atala A (2006). Recent developments in tissue engineering and regenerative medicine. *Current Opinion in Pediatrics*; 18: 167-171.
- De Coppi P, Bartsch G, Siddiqui MM, Xu T, Santos TX and Perin L (2007). Isolation of amniotic stem cell lines with potential for therapy. *Nature Biotechnology*; 25: 100-106.
- Dobreva MP, Pereira PN, Deprest J and Zwijsen A (2010). On the origin of amniotic stem cells: of mice and men. *International Journal of Developmental Biology*; 54: 761-777.
- Dev K, Gautam SK, Giri SK, Kumar A, Yadav A, Verma V, Kumar P, Singh B (2012). Isolation, culturing and characterization of feeder-independent amniotic fluid stem cells in buffalo (*Bubalus bubalis*). *Research in Veterinary Sciences*. 93(2):743-748.
- Fauza D (2004). Amniotic fluid and placental stem cells. *Best Practical Research in Clinical Obstetrics and Gynecology*; 18: 877-891.
- Gosden CM (1983). Amniotic fluid cell types and culture. *British Medical Bulletin*; 39: 348-354.
- Hummon AB, Lim SR, Difilippantonio MJ and Ried T (2007). Isolation and solubilization of proteins after trizol extraction of RNA and DNA from patient material following prolonged storage. *BioTechniques*; 42: 467-470.
- Mihu CM, Ciucu DR, Soritau O, Susman S and Mihiu D (2009). Isolation and characterization of mesenchymal stem cells from the amniotic membrane. *Romanian Journal of Morphology and Embryology*; 50: 73-77.
- Parolini O, Soncini M, Evangelista M and Schmidt D. (2009). Amniotic membrane and amniotic fluid-derived cells: potential tools for regenerative medicine? *Review Regenerative Medicine*; 4: 275-291.
- Sritanaudomchai H, Pavasuthipaisit K, Kitiyanant Y, and Kupradinun P Mitalipov S and Kusamran T (2007). Characterization and multilineage differentiation of embryonic stem cells derived from a buffalo parthenogenetic embryo. *Molecular Reproduction and Development*; 74: 1295-1302.
- Verma V, Gautam SK, Singh B, Manik RS, Palta P, Singla SK, Goswami SL and Chauhan MS (2007). Isolation and characterization of embryonic stem cell-like cells from in vitro-produced buffalo (*Bubalus bubalis*) embryos. *Molecular Reproductive Development*; 74: 520-529.
- Zhao XE and Zheng YM (2010). Development of cloned embryos from porcine neural stem cells and amniotic fluid-derived stem cells. *Animal*; 4: 921-929.
- Zheng YB, Gao ZL, Xie C, Zhu HP, Peng L, Chen JH and Chong YT (2008). Characterization and hepatogenic differentiation of mesenchymal stem cells from human amniotic fluid and human bone marrow: A comparative study. *Cell Biology International*; 32: 1439-1448.